



From gene networks to drugs: systems pharmacology approaches for AUD

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Abstract

The alcohol research field has amassed an impressive number of gene expression datasets spanning key brain areas for addiction, species (humans as well as multiple animal models), and stages in the addiction cycle (binge/intoxication, withdrawal/negative effect, and preoccupation/anticipation). These data have improved our understanding of the molecular adaptations that eventually lead to dysregulation of brain function and the chronic, relapsing disorder of addiction. Identification of new medications to treat alcohol use disorder (AUD) will likely benefit from the integration of genetic, genomic, and behavioral information included in these important datasets. Systems pharmacology considers drug effects as the outcome of the complex network of interactions a drug has rather than a single drug-molecule interaction. Computational strategies based on this principle that integrate gene expression signatures of pharmaceuticals and disease states have shown promise for identifying treatments that ameliorate disease symptoms (called *in silico* gene mapping or connectivity mapping). In this review, we suggest that gene expression profiling for *in silico* mapping is critical to improve drug repurposing and discovery for AUD and other psychiatric illnesses. We highlight studies that successfully apply gene mapping computational approaches to identify or repurpose pharmaceutical treatments for psychiatric illnesses. Furthermore, we address important challenges that must be overcome to maximize the potential of these strategies to translate to the clinic and improve healthcare outcomes.

Keywords Systems pharmacology · Network medicine · *In silico* gene mapping · Connectivity map · L1000 · LINCS · Alcoholism · Alcohol dependence · Gene expression · Transcriptome

Introduction

Developing more effective pharmacotherapies to treat disease is an important goal in public health. This is especially true for complex psychiatric diseases like alcohol use disorder (AUD), where there are limited pharmaceutical treatment options. We use AUD throughout this review for consistency as this is the terminology used in the current edition of the *Diagnostic and Statistical Manual of Mental Disorders (DSM5)*, but this does

not exclude previous DSM version diagnostic criteria or preclinical/clinical trials based on those. AUD is a chronic, relapsing disease that devastates individuals, families, and society and is a major public health problem. Though recovery is possible regardless of disease severity, there are few pharmaceutical treatments available to aid in the recovery process. There are several points of intervention along the time course of AUD where pharmacotherapies might be effective, including AUD initiation (initial alcohol use), development (sporadic intermittent alcohol use; the binge-intoxication phase), progression (regular use), early abstinence (the withdrawal—negative affect stage) or protracted abstinence (the preoccupation—anticipation (craving) stage) (Koob et al. 2009; Kreek et al. 2002). Sleep disturbances are a key contributor to relapse in abstinence and therefore offer another target for treatment (Brower 2015; Miller et al. 2017). Therapeutic interventions at any point along this continuum could improve the health of the individual.

Pharmaceutical treatments can either be developed *de novo* for a specific drug target, repurposed, or rescued. While the

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usage and definition of the terminology “drug repurposing” and “drug rescue” can be complex (Langedijk et al. 2015), here we define drug repurposing as finding a novel clinical use for an approved drug and drug rescue as finding a clinical use for a stalled drug (whether the drug is in development but not yet approved or failed for one indication but could be useful for another disease or patient subgroup; phase 2 or beyond). Drug repurposing (also referred to as drug repositioning) is appealing because it reduces the overall costs of drug development and expedites the availability of treatments to those who need them (Nosengo 2016). Drug repurposing has largely centered around side-effect data, and, while this approach has been somewhat successful for brain diseases, there is a great need for improved strategies for drug selection. De novo drug development has traditionally relied on target identification through basic research. Over four decades of alcohol research has identified key neurotransmitter systems and brain regions that contribute to the various stages of AUD pathology and represent potential targets for pharmaceutical development. Despite these advancements, there has been sparse translational success clinically. There are only three FDA approved drugs for AUD: naltrexone (oral: ReVia®, injectable: Vivitrol®), acamprosate (Campral®), and disulfiram (Antabuse®), the most recent of which, acamprosate, was approved in 2004. This gap between advances in basic research (conducted primarily at academic institutions) and pharmaceutical development (primarily undertaken by industry, e.g., pharmaceutical companies) has been dubbed the “valley of death” (Butler 2008).

The explosion of both the quantity and availability of various types of molecular datasets (e.g., gene sequence/genotype, gene expression, epigenetic marks, metabolic measures) and computational strategies to exploit them, offers new solutions to this problem and is moving disease diagnosis and treatment into the molecular realm. Many computational (or in silico) strategies exist, and all are concerned with finding the “similarity” between diseases and drugs. The computational strategies highlighted in this Review involve integrating molecular profiles of a disease state with those of pharmaceuticals to predict effective treatments. Molecular profiles can be derived from multiple molecular phenotypes, including gene expression, protein targets (see the issue in this article for proteome targets in the accumbens by Clyde Hodge and colleagues), genetic variants (single nucleotide polymorphisms (SNPs)), and others, though the focus of this review will be on gene expression. Another approach to computational repurposing uses crystal structures of receptors to conduct structure-based ligand discovery (Heusser et al. 2013). In this review, we focus on the aforementioned computational approaches and will not discuss structure-based ligand discovery in detail, but interested readers are referred to a review (Howard et al. 2014).

Traditional approach

Drug development

Disease-related drug development begins with mechanistic studies of target identification followed by validation (see the review in this issue by Ciccocioppo and colleagues for an in-depth discussion of target validation), preclinical and clinical trials, and FDA review. Typically, a *single* cellular or molecular target is sourced from the results of much neurobiological research (Fig. 1). Despite clear scientific evidence for its involvement in disease pathology at multiple levels of analysis (e.g., molecular, neuropharmacological, neurocircuitry, behavior), the single target approach has largely been a failure for brain diseases (Hutson et al. 2017). A striking example of this is for Huntington’s disease, where the single causative gene (HTT) has been known since 1993 (MacDonald et al. 1993). Despite this single, well-validated target, no drug nor therapeutic options have been developed as treatments. One example of this for AUD is the corticotrophin releasing factor (CRF) system, which has tremendous research support for its involvement in AUD pathology, yet CRF inhibitors have produced disappointing results in double-blind, placebo-controlled trials (Kwako et al. 2015; Pomrenze et al. 2017; Schwandt et al. 2016).

Despite the vital insights gained from neurobiological research (both in humans and animal models), these findings have not translated into therapeutic success. There are a number of possible reasons for this, including genetically heterogeneous human populations and the complexities of alcohol’s many targets (Most et al. 2014; Pomrenze et al. 2017). The brain is highly complex, and psychiatric diseases are characterized by numerous symptoms. Reducing this complexity to a single target is appealing for its simplicity but perhaps misguided, and expecting modulation of a single gene or molecule to ameliorate all symptoms of complex diseases is likely to produce disappointing results.

Targets (molecules) do not work in isolation, but function as part of a system (or network) to accomplish biological functions. The hypothesis that a disease state represents a shift from normal physiological homeostasis and can be thought of as a network perturbation has been proposed and described in detail, and is attractive for several reasons (Barabasi et al. 2011; Chen and Butte 2013; Jacunski and Tatonetti 2013; Kolodkin et al. 2012; Silbersweig and Loscalzo 2017; Silverman and Loscalzo 2013). First, there could be many network perturbations that lead to the same disease classification, which fits with the heterogeneous patient populations we observe in AUD. Secondly, the other side of this argument is that if a disease represents a perturbed state of a biological network, there could be multiple pharmacological intervention points to reverse those perturbations and return the system to homeostasis. Targeting the network at several points might

Traditional Approach

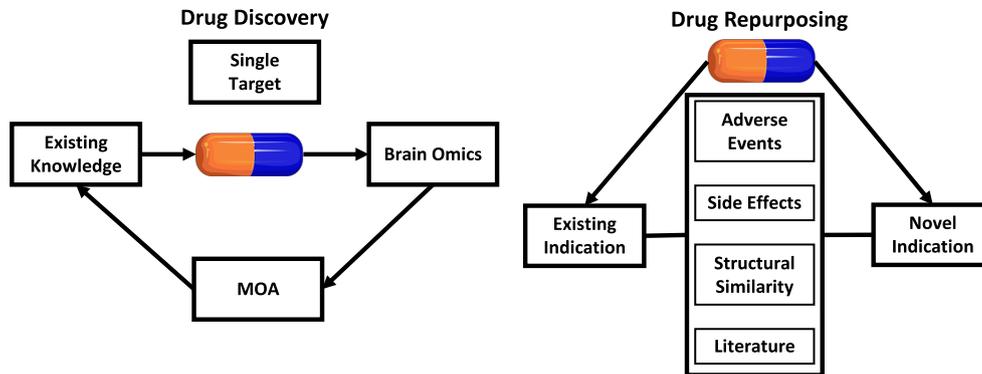


Fig. 1 Traditional approach to drug discovery and drug repurposing: existing knowledge of a disease state (built upon basic science) is applied to select a compound designed for a single target (chosen for its involvement in a disease process), and these are tested in vitro and/or in vivo. Brain gene expression levels (Brain Omics) are measured for drugs that ameliorate disease phenotype which helps further elucidate the mechanisms of action (MOAs) of drugs and suggests other molecules that can be targeted by candidate drugs. Traditionally, drug repurposing (finding new indications for existing compounds) has been largely based

on side effect data, adverse events, existing literature, or structural similarity between compounds used to treat different diseases (the idea being that the compound of one disease might be able to treat another because it shares structural similarity with compounds used to treat that disease). Drug repurposing efforts would benefit greatly if there was a system established to report positive side effects as is the case for “adverse events.” Capsule images from <http://smart.servier.com/category/general-items/drugs-and-treatments/>. Servier Medical Art by Servier is licensed under CC BY 3.0 (<https://creativecommons.org/licenses/by/3.0/>)

be more efficient (or even necessary) to shift the system back to normal homeostasis. This also provides a basis for polypharmacology (the use of drug combinations to treat a disease) and could guide the selection of drug combinations, which will not be discussed in depth in this review, but interested readers are referred to Ryall and Tan (2015) for more information. For these reasons, we and others propose that to maximize the likelihood of successful treatment for complex disorders, it is imperative to “drug the network” rather than focus solely on single targets (see “Computational approaches” section).

Drug repurposing

Traditionally, getting a drug to market takes 13–15 years and costs 2–3 billion dollars on average (Nosengo 2016). Many drugs that are currently FDA approved could be beneficial for diseases other than their original indication. Additionally, pharmaceutical companies have invested considerable resources into developing drugs that passed initial safety trials but failed in efficacy trials (sometimes referred to as shelved compounds) that are waiting for a suitable indication (Nosengo 2016). Often, successful drug repurposing has been serendipitous (Fig. 1). There are many examples spanning a variety of conditions, from the classic example of sildenafil (Viagra®), a PDE5 inhibitor being developed for hypertension, that was repurposed for erectile dysfunction (Goldstein et al. 1998), to bimatoprost (Lumigan®/Latisse®), a prostaglandin analog that was repurposed for a cosmetic application as it was

noticed to lengthen and darken the eyelashes as a side effect of those using it to treat glaucoma (Tosti et al. 2004).

Drug repurposing has also been successful for brain diseases. For example, buprenorphine, a mixed partial agonist opioid receptor modulator, was originally used for pain relief and was repurposed to treat opiate dependence (Jasinski et al. 1978). Ropinirole (Requip), a dopamine agonist used as an anti-Parkinson’s agent, was repurposed for treatment of both restless legs syndrome and SSRI-induced sexual dysfunction (Cheer et al. 2004; Worthington et al. 2002). Additional examples include bupropion (depression to smoking cessation) (Lief 1996), dimethyl fumarate (psoriasis to multiple sclerosis) (Bomprezzi 2015), and guanfacine (hypertension to ADHD) (Strange 2008).

Several FDA approved or shelved compounds have shown promise in treating AUD and many are currently undergoing human lab testing or are in clinical trials (ClinicalTrials.gov_AUD), including gabapentin, topiramate, varenicline, ABT-436, mifepristone (RU-486), citicoline, baclofen, nalmefene, and others (Litten et al. 2016; Lyon 2017) (Table 1).

Gabapentin (Neurontin) was initially used as an anti-epileptic, then later approved for neuropathic pain and amyotrophic lateral sclerosis. Baclofen (Lioresal) is a GABA_B receptor agonist, originally made as an anti-epileptic with disappointing results, but showed remarkable effectiveness for treating spasticity in many conditions, especially for spinal cord injury, cerebral palsy, and multiple sclerosis. As mentioned, it is being considered for treatment of AUD (with mixed findings) (Farokhnia et al. 2017).

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Table 1 Drugs with potential to be repurposed for AUD

Drug name	Target	Original indication	Novel indication	Clinical trial	Refs
Quetiapine (Seroquel)	Atypical antipsychotic. Antagonist: D1 and D2 dopamine receptors, alpha 1 and alpha 2 adrenoreceptors, and 5-HT1A and 5-HT2 serotonin receptors, histamine H1 receptor (and others)	Schizophrenia (Sez) Bipolar disorder (BD). Major depressive disorder (MDD) (along with an SSRI)	AUD and bipolar; sleep disturbances in abstinence; schizophrenia and AUD and anxiety	NCT00457197, NCT00114686, NCT00223249, NCT00550394, NCT00434876, NCT00156715, and NCT00352469	Jensen et al. (2008), Schotte et al. (1996), and Litten et al. (2012)
Aripiprazole (Abilify)	Atypical antipsychotic. Antagonist: D1 and D2 dopamine receptors, alpha 1 and alpha 2 adrenoreceptors, and 5-HT1A and 5-HT2 serotonin receptors, histamine H1 receptor (and others)	Sez, BD, and MDD (along with an SSRI)	AUD and bipolar	NCT02918370	Anton et al. (2008), Kema et al. (2009), Martinotti et al. (2009), Martinotti et al. (2007), Shapiro et al. (2003), and Voronin et al. (2008)
Duloxetine (Cymbalta)	Serotonin–norepinephrine reuptake inhibitor (SNRI)	MDD, GAD, muscle pain, and peripheral neuropathy	AUD	NCT00929344	Bymaster et al. (2001)
Venlafaxine (Effexor)	SNRI	MDD, GAD, panic disorder, and social anxiety disorder	AUD and anxiety	NCT00248612	Bymaster et al. (2001), Ciraulo et al. (2013), and Upadhyaya et al. (2001)
Rolipram	Phosphodiesterase-4 inhibitor	Shelved compound; phase 3 for MDD (Fleischhacker et al. 1992)	AUD		Bell et al. (2017), Dominguez et al. (2016), Gong et al. (2017), Hu et al. (2011), Liu et al. (2017), Ray et al. (2014), and Wen et al. (2012)
Ibudilast	Phosphodiesterase inhibitor	Asthma (Japan)	AUD	NCT02025998	Bell et al. (2015), Crews et al. (2017), Ray et al. (2017), and Ray et al. (2014)
Fenofibrate (Tricor)	Fibrate, PPAR α agonist	Hypercholesterolemia and hypertriglyceridemia	AUD	NCT02158273	Blednov et al. (2015), Blednov et al. (2016a, b), Ferguson et al. (2014), Haile and Kosten (2017), Karahanian et al. (2014), and Rivera-Meza et al. (2017)
Gabapentin (Neurontin)	Anticonvulsant binds to the $\alpha 2\delta$ subunit of the voltage-dependent calcium channels (Pregabalin is structurally related to gabapentin)	Seizures, restless leg syndrome, postherpetic neuralgia, shingles	AUD; abstinence initiation in AUD; AUD (in combination with naltrexone); Sleep disturbances in AUD; AUD (in combination with flumazenil for withdrawal and relapse prevention); AUD (in combination	NCT02771925, NCT01141049, NCT00391716, NCT00183196, NCT01014533, NCT00262639, NCT03274167, NCT00011297, NCT03205423, and NCT02252536	Geisler and Ghosh (2014), Guglielmo et al. (2012), Litten et al. (2016), Mason et al. (2014a, b), and Nunes (2014)

Table 1 (continued)

Drug name	Target	Original indication	Novel indication	Clinical trial	Refs
Pregabalin (Lyrica)	Anticonvulsant binds to the $\alpha 2\delta$ subunit of the voltage-dependent calcium channels	Epilepsy, neuropathic pain, fibromyalgia, generalized anxiety disorder (GAD)	with lorazepam for withdrawal); comorbid alcohol and opioid abuse AUD; AUD and PTSD	NCT03256253, NCT02884908, and NCT00929344	Guglielmo et al. (2012) and Li et al. (2011)
Topiramate (Topamax)	Anticonvulsant blocks voltage-gated Na^+ channels, PAMs of subunits of the GABA_A receptor, modulates AMPA/kainite glutamate receptors, and blocks carbonic anhydrase (CA) CA II and CA IV	Epilepsy, migraines	AUD; AUD and PTSD; AUD and Borderline Personality Disorder; AUD and BD; AUD and cocaine dependence; AUD and nicotine dependence	NCT01135602, NCT01145677, NCT01749215, NCT00769158, NCT00463775, NCT00210925, NCT00572117, NCT00223639, NCT00884884, NCT00006205, NCT00571246, and NCT03120468 NCT00802412 NCT00448825 NCT00329407 NCT00862563 NCT01182766 NCT00300742 NCT00167245 NCT02371889 NCT01764685 NCT01087736 NCT01408641 NCT00550394 NCT03018704	Guglielmo et al. (2015), Ray and Bujarski (2016), and Shank et al. (2000)
Varenicline (Chantix and Champix)	$\alpha 7$ nicotinic acetylcholine receptor agonist; $\alpha 4\beta 2$, $\alpha 3\beta 4$, and $\alpha 6\beta 2$ subtypes partial agonist; weak agonist on the $\alpha 3\beta 2$ -containing receptors	Smoking cessation	AUD; AUD and tobacco dependence; AUD and cocaine dependence; AUD, SCZ, and nicotine dependence	NCT01071187, NCT01146613, NCT00705523, NCT00846859, NCT00873535, NCT01553136, NCT01347112, NCT01151813, NCT01169610, NCT00727103, NCT01011907, NCT01092702, NCT01286584, NCT01592695, and NCT02698215 NCT01613014	Falk et al. (2015) and Litten et al. (2013)
ABT-436	Highly selective vasopressin V1B receptor antagonist	Shelved compound; phase 2 for MDD (NCT01741142)	AUD		Ryan et al. (2017)
Mifepristone (RU-486) (Mifeprex)	Glucocorticoid and progesterone receptor antagonist	Abortifacient, hyperglycemia, and diabetes mellitus	AUD	NCT02243709, NCT02179749, NCT02989662, and NCT01548417	Donoghue et al. (2016), Howland (2013), Lyon (2017), Vendruscolo et al. (2012), and Vendruscolo et al. (2015)
			AUD; AUD and BD	NCT02074735 and NCT02582905	

Table 1 (continued)

Drug name	Target	Original indication	Novel indication	Clinical trial	Refs
Citicoline (Cebroton, Ceraxon, Cidilin, Citifar, Cognizin, more)	Membrane permeability enhancer. Glutathione transferase stimulant (over-the-counter nutritional supplement)	Stroke, Alzheimer's disease, senile dementia, Parkinson's disease, attention-deficit/hyperactivity disorder (ADHD), and glaucoma	Stroke, Alzheimer's disease, senile dementia, Parkinson's disease, attention-deficit/hyperactivity disorder (ADHD), and glaucoma		Secades and Lorenzo (2006) and Wignall and Brown (2014)
Baclofen (Lioresal)	Central nervous system depressant; skeletal muscle relaxant. GABA _B receptor agonist	Spastic movement disorders (commonly for spinal cord injury, cerebral palsy, and multiple sclerosis)	AUD; alcohol withdrawal; AUD and Hepatitis C; AUD with liver disease; AUD and anxiety disorders	NCT02596763, NCT03034408, NCT03293017, NCT01008280, NCT02511886, NCT01711125, NCT01751386, NCT00877734, NCT00614328, NCT01266655, NCT01980706, NCT01738282, NCT01002105, NCT00802035, NCT02835365, NCT01604330, NCT02723383, NCT01076283, NCT01937364, NCT00525252, NCT02107352, and NCT02771925	Bell et al. (2017), Borro et al. (2016), Colombo et al. (2004), Farokhnia et al. (2017), Geisel et al. (2016), Imbert et al. (2015), Litten et al. (2016), Liu and Wang (2017), Lyon (2017), Mirijello et al. (2015), Morley et al. (2014), Muller et al. (2015), Pomizovsky et al. (2015), Rigal et al. (2015), Rolland et al. (2015a, b), and Weibel et al. (2015)
Nalmefene (Selincro)	Antagonist of the μ -opioid receptor; weak partial agonist of the κ -opioid receptor	Antidote for opioid overdose. Approved in Europe for AUD	AUD; AUD with cirrhosis; AUD and tobacco dependence; AUD and borderline personality disorder; AUD and opioid use disorder	NCT00811720, NCT01969617, NCT00812461, NCT00811941, NCT02824354, NCT02382276, NCT02364947, NCT02197598, NCT02679469, NCT02372318, NCT02195817, NCT02492581, NCT00000450, NCT00000437, NCT02752503, NCT03034408, and NCT03279562	Litten et al. (2016), Naudet (2016), Naudet et al. (2016), Soyka (2016), and Soyka et al. (2016)

Computational approaches

The generation and accumulation of publicly accessible, high-throughput genomic datasets make it possible to integrate large-scale drug and disease signatures at the molecular level to predict compounds with the potential to treat a disease based on multiple targets (e.g., gene networks). These data-rich resources include public repositories (primary archives), integrative databases, and value-added databases (these tools are designed to process, analyze and annotate complex information from primary data sources to lower the computational barriers to access primary data). A selection of these resources is summarized in Table 2.

There are two essential datasets from these resources that are required to match disease and drug: (1) measurements of a molecular phenotype induced by a disease state and (2) measurements of the same molecular phenotype induced by drugs. Obtaining this type of reliable drug library is not trivial. Surprisingly, attaining a list of approved drugs and their indications is not a straightforward task. These difficulties are the result of poor data storage and electronic retrieval mechanisms, complex and rapidly changing nomenclature (drugs can be called by their common name, chemical name, simplified molecular-input line-entry system (SMILES), International Chemical Identifier (InChI)), and legal issues surrounding off-label advertisement of pharmaceuticals. Fortunately, many of these challenges have been overcome largely by the pioneering work by a collaborative effort of the Broad Institute and funding from a National Institutes of Health Common Fund (<https://commonfund.nih.gov/lincs>). They have compiled the Library of Integrated Network-based Cellular Signatures (LINCS-L1000) database which contains gene expression responses to genetic and pharmacologic manipulation across a diverse set of human cell lines (Subramanian et al. 2017). They also maintain a repurposing hub that contains over 5000 manually-curated drugs that are either FDA approved or in clinical trials (Corsello et al. 2017). The availability of these tremendous resources is a primary reason we focus on gene expression as the molecular phenotype, as other molecular responses to drugs are not as well characterized in such a systematic manner or as accessible for analysis. With the two required datasets described above, there are three main steps to proceed from gene networks to candidate compounds (see Fig. 2), which then can be tested in a preclinical animal model or human laboratory study:

1. Generate an input signature that captures the genomic state of interest (gene expression differences between disease and healthy state, for example).

2. High-throughput identification of compounds using an in silico screen (similarity metric).
3. Prioritize candidate compounds.

The details of each step are described below.

Generate an input signature that captures the genomic state of interest

The purpose of the signature is to capture the molecular changes that are the most relevant to the biological state of interest at a given point in time. There are many different options for constructing an input signature. Applying such approaches to brain diseases is still in its infancy and understanding the optimal input parameters is a major challenge (see the “Challenges and future directions” section below). Genetic variation (genotyping or exome/whole genome sequencing data) has been the primary approach used for genomic medicine/precision medicine for cancer (Letai 2017). A functional genomic measure, such as gene expression can also be used. This is referred to as in silico gene mapping, gene mapping, or connectivity mapping, the latter named after one of the first characterizations of the method using the Broad Institute’s database called the Connectivity Map (CMap) (Lamb et al. 2006). Importantly, the AUD research field has generated an incredible library of gene expression data that spans multiple species (human, mouse, monkey, rat), conditions/treatments (genetic predisposition, various acute or chronic ethanol exposures, or paradigms), various brain regions, and isolated cell types (including microglia and astrocytes) (Table 2).

High-throughput identification of compounds using an in silico screen (similarity metric)

At their core, the various approaches used for in silico gene mapping aim to compare drug and disease signatures. If an effect size measure (such as fold change) is available, a correlation coefficient could be calculated, to reflect the correlation between gene expression changes between drug and vehicle and those between disease and normal. Positive correlations would indicate that the drug mimics the disease’s effects on transcription levels, while negative correlations would indicate that the drug reverses it. An alternative approach is to use an enrichment score to assess the overlap between two lists of differentially expressed genes, such as the hypergeometric statistic or the rank-based gene set enrichment analysis (GSEA; corresponds to a weighted Kolmogorov-Smirnov) (Subramanian et al. 2005). For example, list A contains the top differentially expressed genes between disease and healthy samples, and list B contains the top differentially expressed genes between drug and vehicle samples. The

Table 2 Data resources

Name	Description	URL	Ref.
Primary repositories			
Gene Expression Omnibus	Public functional genomics data repository for array- and sequence-based data.	ncbi.nlm.nih.gov/geo/	Barrett et al. (2013)
ArrayExpress	Public functional genomics data repository for array- and sequence-based data.	ebi.ac.uk/arrayexpress/	Kolesnikov et al. (2015)
ParkDB	Repository for gene expression datasets related to Parkinson's disease (PD)	www2.cancer.ucl.ac.uk/Parkinson_Db2/	Taccioli et al. (2011)
Integrative databases			
HUGO Gene Nomenclature Committee (HGNC) database	Repository of HGNC-approved gene nomenclature, gene families and associated resources including links to genomic, proteomic, and phenotypic information.	genenames.org/	Gray et al. (2015)
Online Mendelian Inheritance in Man (OMIM)	Publicly available dataset of human genes and genetic disorders and traits, with particular focus on the molecular relationship between genetic variation and phenotypic expression.	omim.org/	Amberger and Hamosh (2017)
UK Brain Expression Consortium	Publicly available dataset of genotyping and gene expression data from 134 brains from individuals free of neurodegenerative disorders (up to 12 brain regions).	ukbec.wordpress.com/braineac.org/	
Encyclopedia of DNA Elements (ENCODE)	Integrates multiple technologies and approaches in a collective effort to discover and define the functional elements encoded in the human genome, including genes, transcripts, and transcriptional regulatory regions, together with their attendant chromatin states and DNA methylation patterns.	encodeproject.org/	Consortium (2011)
Genotype–Tissue Expression (GTEx) project	Collection and analysis of multiple human tissues from donors who are also densely genotyped, to assess genetic variation within their genomes. By analyzing global RNA expression within individual tissues and treating the expression levels of genes as quantitative traits, variations in gene expression that are highly correlated with genetic variation can be identified as expression quantitative trait loci, or eQTLs.	gtexportal.org/home/	Consortium (2013)
Depression Genes and Networks (DGN) cohort	RNA sequencing data and analyses from 922 genotyped individuals, providing information regarding the regulatory consequences of genetic variation	dags.stanford.edu/dgn/	Battle et al. (2014)
Psychiatric Genomics Consortium (PGC)	Psychiatric Genomics Consortium (PGC) unites investigators around the world to conduct meta- and mega-analyses of genome-wide genomic data for psychiatric disorders.	med.unc.edu/pgc	O'Donovan (2015)
Library of Integrated Network-Based Cellular Signatures; LINCS-L1000	There are samples from more than 900,000 individuals (and growing) collected by over 800 investigators from 38 countries. Publicly available dataset from the Broad Institute. The 1.3 M L1000 cellular signatures catalog transcriptional responses of human cells to chemical and genetic perturbation. A total of 27,927 perturbagens have been profiled to produce 476,251 expression signatures. About half of those signatures make up the Touchstone (reference) dataset generated from testing well-annotated genetic and small-molecular perturbagens in a core panel of cell lines.	clue.io/	Subramanian et al. (2017)
Connectivity Map (CMap)	Publicly available dataset from the Broad Institute. Connectivity Map Build 02 includes data from 7056 Affymetrix microarrays, for 1309 small-molecule compounds, and 6100 treatment instances in 5 human cell lines.	broadinstitute.org/cmap/	Lamb et al. (2006)
Added-value databases and tools			
Genes Enrichr	Web tool for gene set enrichment analysis	amp.pharm.mssm.edu/Enrichr/	Kuleshov et al. (2016)
NetworkAnalyst	Web tool for performing various common and complex meta-analyses of gene expression data	networkanalyst.ca/	Xia et al. (2015)
Database for Annotation, Visualization and Integrated Discovery (DAVID)	Web tool for gene set enrichment analysis	david.ncifcrf.gov/	Huang et al. (2009)
GeneMANIA	Web tool for generating hypotheses about gene function, analyzing gene lists and prioritizing genes	genemania.org/	Montejo et al. (2014)

Table 2 (continued)

Name	Description	URL	Ref.
WebGestalt (WEB-based Gene Set Analysis Toolkit)	for functional assays. There is also a GeneMANIA Cytoscape plugin. Web tool for gene set enrichment analysis.	webgestalt.org	Wang et al. (2013)
PubMatrix	Web tool that allows simple text based mining of the NCBI literature search service PubMed using any two lists of keywords terms, resulting in a frequency matrix of term co-occurrence.	pubmatrix.irp.nia.nih.gov/	Becker et al. (2003)
Ingenuity Pathway Analysis (IPA®)	Tool for analyzing and visualizing data from omics experiments	qiagenbioinformatics.com/products/ingenuity-pathway-analysis/	Kramer et al. (2014)
Gene-Set Enrichment Analysis (GSEA)	Web tool for determining whether an a priori-defined set of genes shows statistically significant, concordant differences between two biological states (phenotypes).	broadinstitute.org/gsea/	Subramanian et al. (2005)
MetaXcan	Algorithm that allows imputation of gene expression z-scores based on GWAS summary statistics.	github.com/hakyimlab/MetaXcan	Barbeira et al. (2016)
Kyoto Encyclopedia of Genes and Genomes (KEGG)	Database resource for understanding high-level functions and utilities of the biological system, such as the cell, the organism and the ecosystem, from molecular-level information, especially large-scale molecular datasets generated by genome sequencing and other high-throughput experimental technologies. (So et al. 2017) used KEGG to download the Anatomical Therapeutic Classification (ATC) for drugs.	genome.jp/kegg/	Ogata et al. (1999)
GeneGO's Metacore	Integrated software suite for functional analysis of experimental data based on a curated database of human protein-protein, protein-DNA interactions, transcription factors, signaling and metabolic pathways, disease and toxicity, and the effects of bioactive molecules. Suite contains tools for data visualization, mapping and exchange, multiple networking algorithms, and filters.	portal.genego.com/	Ekins et al. (2006)
GeneWeaver	Curated repository of genomic experimental results from published genome-wide association studies, quantitative trait locus, microarray, RNA-sequencing and mutant phenotyping studies with an accompanying tool set for dynamic integration of these data sets, enabling users to identify gene-function associations across diverse experiments, species, conditions, behaviors, or biological processes.	geneweaver.org/	Baker et al. (2012)
GeneCards	Database of human genes that provides genomic, proteomic, transcriptomic, genetic, and functional information on all known and predicted human genes. Developed and maintained by the Crown Human Genome Center at the Weizmann Institute of Science.	genecards.org	Stelzer et al. (2016)
TransFind	Web tool for predicting transcriptional regulators for gene sets	transfind.sys-bio.net/	Kielbasa et al. (2010)
JASPAR	Web tool for predicting transcriptional regulators for gene sets	jaspar.genereg.net/	Mathelier et al. (2016)
TRANSFAC (TRANScription FACTor database)	Web tool for predicting transcriptional regulators for gene sets	gene-regulation.com/pub/databases.html	Matys et al. (2006)
Proteins STRING	Web tool/database that provides a critical assessment and integration of protein-protein interactions, including direct (physical) as well as indirect (functional) associations.	string-db.org	Szklarczyk et al. (2015)
iRefWeb	Web tool/database that integrates data on protein-protein interactions (PPI) consolidated from major public databases.	wodaklab.org/iRefWeb/	Turinsky et al. (2014)
Hippie	Web tool to generate reliable and meaningful human protein-protein interaction networks.	cbdm-01.zdv.uni-mainz.de/~mschaefer/hippie/	Alanis-Lobato et al. (2017)
Drugs sscMap	Java application that performs connectivity mapping tasks using the CMap build 02 data. Users can add custom collections of reference profiles.	purl.oclc.org/NET/sscMap	Zhang and Gant (2009)
Searchable Platform Independent Expression Database Webtool (SPIEDw)	Web tool used for querying publically available gene expression data (including the CMap build 02 drug data).	spied.org.uk/cgi-bin/HGNC-SPIED3.1.cgi	Williams (2012)
Drug-Set Enrichment Analysis (DSEA)	Web tool for identifying shared pathways whose genes are upregulated (or downregulated) by the drugs in the set.	dsea.tigem.it/	Napolitano et al. (2016)

Table 2 (continued)

Name	Description	URL	Ref.
ChemBioServer	Web tool for mining and filtering chemical compounds used in drug discovery	bioserver-3.bioacademy.gr/Bioserver/ChemBioServer/	Athanasiadis et al. (2012)
Mode of Action by NeTwoRk Analysis (Mantra 2.0)	Web tool for the analysis of the Mode of Action (MoA) of novel drugs and the identification of known and approved candidates for “drug repositioning” using CMap drug data.	mantra.tigem.it/	Carrella et al. (2014)
Comparative Toxicogenomics Database (CTD)	Publicly available dataset describing relationships between chemicals, genes, and human diseases.	ctdbase.org/	Davis et al. (2017)
MEDication Indication resource (MEDI)	Compiled from four public medication resources, including RxNorm, Side Effect Resource 2 (SIDER2), Wikipedia and MedlinePlus. A random subset of the extracted indications was also reviewed by physicians. The MEDI high-precision subset (MEDI-HPS), only includes drug indications found in RxNorm or in at least two of the other three sources, with an estimated precision of 92%.	vumc.org/cpm/center-precision-medicine-blog/mediensemble-medication-indication-resource	Wei et al. (2013)
ClinicalTrials.gov National Institute for Occupational Safety and Health List of Antineoplastic and Other Hazardous Drugs DrugBank	Contains information about clinical trials. Contains drugs known to be toxic, according to published literature	ClinicalTrials.gov cdc.gov/niosh/topics/hazdrug/default.html	Traynor (2014)
DrugBank	Bioinformatics and cheminformatics resource that combines detailed drug data with comprehensive drug target information.	DrugBank.ca/	Wishart et al. (200)
STITCH	Database that includes information on chemical-protein interactions. The interactions include direct (physical) and indirect (functional) associations; they stem from computational prediction, from knowledge transfer between organisms, and from interactions aggregated from other (primary) databases. Currently it has 9,643,763 proteins from 2031 organisms.	stitch.embl.de/	Szklarczyk et al. (2016)
PharmGKB	Repository for pharmacogenetic and pharmacogenomic data, and curators provide integrated knowledge in terms of gene summaries, pathways, and annotated literature.	pharmgkb.org	Owen et al. (2007)
SuperTarget	Added-value database that integrates information about drugs, proteins and side effects from other databases to form drug-protein, protein-protein and drug-side-effect relationships and includes annotation about the source, ID's, physical properties, references and much more.	insilico.charite.de/supertarget/	Hecker et al. (2012)
KEGG Drug	Comprehensive drug information resource for approved drugs in Japan, USA, and Europe unified based on the chemical structures and/or the chemical components, and associated with target, metabolizing enzyme, and other molecular interaction network information.	genome.jp/kegg/drug/	Ogata et al. (1999)
AUD specific INIA Texas Gene Expression Database (IT-GED)	Contains the top statistical results from genomic studies focusing on models of excessive alcohol consumption.	inia.icmb.utexas.edu/	
Ethanol-Related Gene Resource (ERGR)	Contains more than 30 large datasets from literature and 21 mouse QTLs from public database (see data summary). These data are from 5 organisms (human, mouse, rat, fly, and worm) and produced by multiple approaches (expression, association, linkage, QTL, literature search, etc.)	bioinfo.uth.edu/ERGR/	Guo et al. (2009)
Gene Network	Contains large collections of genotypes (e.g., SNPs) and phenotypes that are obtained from groups of related individuals, including human families, experimental crosses of strains of mice and rats, and organisms as diverse as <i>Drosophila melanogaster</i> , <i>Arabidopsis thaliana</i> , and barley.	genenetwork.org/webqtl/main.py	Mulligan et al. (2017)

hypergeometric statistic would give the probability of the overlap between list A and list B (the genes changed by both drug and disease). GSEA, the approach implemented by the

Connectivity Map (CMap) and LINCS-L1000, avoids using arbitrary cutoffs (the p value which designates differential expressions between two conditions or treatments) by

Computational Approach Multiple Targets

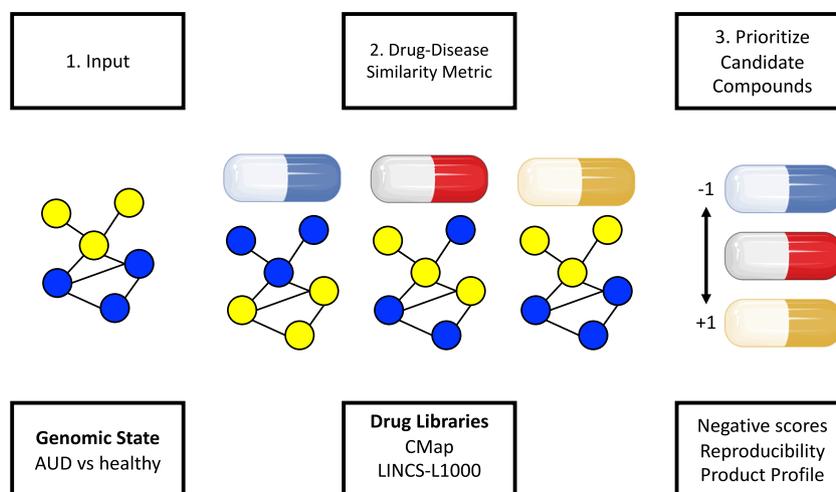


Fig. 2 Computational approach to drug discovery and drug repurposing: disease state can be either acquired (disease or substance of abuse changes gene networks and these changes drive disease) or predisposed (genetic variants cause disruptions in gene networks). The goal of in silico gene mapping is to integrate the targets (gene networks) of disease and drugs to find a drug (or combination of drugs) that affect similar targets as the disease. Drugs that oppose the disease-state's molecular disruption (many targets) are chosen as candidate compounds to ameliorate disease phenotype. There are three steps to go from gene expression datasets to candidate compounds: (1) generate an input genomic signature or network. Shown is a gene-gene coexpression network of genes related to a disease state: nodes = genes, edges = gene-gene expression correlation, yellow = up-regulated genes, blue = down-regulated genes; (2) compare the disease signature to those induced by drugs to identify drugs that would reverse the disease signature. Shown are the effects of three

different drugs in the reference database (e.g., LINCS-L1000) on the disease-related genes that served as the input and (3) prioritize candidate compounds for in vivo testing. The blue drug that received a perfect negative score would be prioritized because it down-regulated the genes that were up-regulated in the disease state and up-regulated the genes that were down-regulated in the disease state. The yellow drug would be predicted to mimic or worsen the disease state. Had the input been a desirable biological state (e.g., the gene expression profile of patients with AUD who had prolonged recovery vs. those who relapsed quickly after ceasing alcohol consumption), then the yellow drug would be prioritized because it is predicted to mimic the beneficial biological state. Capsule images from <http://smart.servier.com/category/general-items/drugs-and-treatments/>. Servier Medical Art by Servier is licensed under [CC BY 3.0](https://creativecommons.org/licenses/by/3.0/) (<https://creativecommons.org/licenses/by/3.0/>). AUD, alcohol use disorder

considering all of the genes in an experiment. Ranked methods for the hypergeometric test have also been described and offer the same benefits as GSEA (Plaisier et al. 2010).

Prioritize candidate compounds

Regardless of which statistical test is chosen, the output of the previous step will include a list of predicted compounds with a corresponding similarity score (also called a connectivity score). Because only a handful of drugs can be tested in vivo, this list must be filtered to select the most promising candidate compounds. The working hypothesis is that negative scores would predict reversal of gene expression from disease back to normal state. However, this hypothesis is rarely tested directly (see “Challenges and future directions”), though it does have some support (Chen et al. 2017; Delahaye-Duriez et al. 2016; Wagner et al. 2015). Regardless, drugs with either the highest absolute value or the most negative similarity scores should be prioritized, as these reflect the drugs that affect the most disease-related genes.

Beyond the sign (+/−) and magnitude of the connectivity score, there are additional practical considerations for prioritizing candidate compounds (Oprea and Overington 2015). For example, any identified high-priority candidate drugs for AUD treatment would also benefit from having (1) known oral dosing data available, (2) have little or no safety warnings (especially regarding liver toxicity), (3) have low abuse liability, (4) low drug-drug interaction potential, (5) negligible cytotoxic actions, and (6) high brain penetrability, among others. These considerations alone will assist in narrowing the pool of potentially “testable” compounds considerably, if the information is available (which is frequently not the case). Upon first glance, it might seem that the challenge is selecting only a few compounds from hundreds of candidates generated by in silico screens to test clinically or preclinically. However, this is not the case. Meeting the ideal practical considerations outlined above could eliminate virtually all candidate compounds (Oprea and Overington 2015). In that case, medicinal chemistry approaches could be used to modify the chemical structure to suit the desired product profile.

Drug prioritization should also depend on the reliability of the analytical results. That is, the connectivity score should be reproducible for strong candidates. This is especially important to consider because small changes in the genes that makeup the input signature can result in the identification of different candidate compounds. The above-mentioned CMap database utilizes a statistical measure of reliability (permutation test) to achieve this goal. Stricter statistical measures have also been developed for CMap. For example, the statistically significant connectivity map (ssCMap) was developed (Zhang and Gant 2009), which includes a measure of stability by removing single genes from the input in a systematic manner and assessing reproducibility (McArt and Zhang 2011). However, for larger datasets, such as LINCS-L1000, implementation of permutations tests becomes computationally expensive and less straightforward. Currently, the web app for querying the LINCS-L1000 data (<https://clue.io/l1000-query>) uses the “sig_gutc” tool (Subramanian et al. 2017) to summarize the connectivity scores and provide a measure of reliability. Each compound has been profiled under multiple experimental conditions (different cell lines, drug doses, and exposure time points). To attain a compound-level analysis, sig_gutc reports a summary score of the distribution of scores for a compound across all experiments. The tool then ranks the connectivity score between the query signature and the compound signature, based upon the compound’s precomputed distribution of connectivity scores with the other hundreds of thousands of signatures in the LINCS-L1000 database. This provides a measure of the likelihood of a connectivity score for a drug given that drug’s connectivity with the database as a whole, thus mitigating false positives from drugs with widespread effects on transcription. However, an appropriate statistical framework with which to interpret LINCS-L1000 results needs to be developed.

Application to brain diseases

While initially used in cancer research (for review, see Chen and Butte 2016), these computational repurposing strategies have also been applied to brain diseases, albeit in a more limited capacity. However, it should be noted that although not used as widely, the studies using these computational approaches for drug discovery for brain diseases have provided promising leads for variety of disease states. Because there are few applications so far for psychiatric disorders, this review includes the use of *in silico* gene mapping strategies for any disease in which brain is the primary affected organ, for which there have been 20 studies so far to the best of our knowledge (Table 3).

Regarding the construction of the input genomic signature, the studies fall into two main categories: those that use genotype data (i.e., SNPs related to a disease phenotype discovered from genome-wide association studies (GWAS)) and those that use gene expression data. Gene expression measurement technology (RNA sequencing or microarray) provides the expression levels of all genes in the genome simultaneously, supplying a functional

genomic readout of the effects of the combination of the genetic variants that could be contributing to disease. Gene expression is by far the primary input used by the studies in Table 3, the idea being to compare the gene expression levels between disease and healthy tissue and to use the top differentially expressed genes as the input signature, as this is thought to best capture the molecular differences driving disease phenotypes. However, there is no consensus on the optimal threshold or number of differentially expressed genes to use. Differentially expressed genes can be subdivided into groups of genes with highly correlated expression levels. Indeed, several studies incorporate gene co-expression networks or protein-protein interaction networks to refine the genomic input signature (Chandran et al. 2016; Delahaye-Duriez et al. 2016; Gao et al. 2014). One study compared the performance of using only differentially expressed genes between Parkinson’s and normal brain to query CMap versus a combined approach that used both differential expression and gene co-expression network information (Gao et al. 2014). They calculated the number of known Parkinson’s therapeutics in the top 50 ranked compounds from each approach. Using the top 20 genes from the combined method outperformed using the top 20 genes from differential expression alone. They were not able to assess the performance of using only the gene co-expression network as a query for lack of up-regulated genes in the co-expression module. Interestingly, using more than the top 20 genes from the combined approach led to a decrease in performance. Because gene co-expression network modules are driven by variability in the data, and cell type is a major contributor to gene expression variability, it is possible that network based approaches could be more useful for diseases that primarily affect a specific cell type (like in the case for Parkinson’s disease). One downside of using gene expression data is that human brain tissue can only be obtained postmortem and the transcriptional signature can be confounded by a lifetime with the disease or pharmaceutical management of the disease (see “Challenges and future directions”).

Genotype/gene sequence data, on the other hand, is readily available, easy to attain, and is relatively static throughout the patient’s lifetime, but it is not without its drawbacks. Many genes contribute to the genetic risk of most complex psychiatric disorders, each contributing a small effect. A minority of disease-associated SNPs are mapped to protein-coding regions of the genome, and there are few drugs that specifically target particular gene products. Despite these challenges, Papassotiropoulos et al. (2013) used intragenic SNPs related to aversive memory performance to select the antihistamine, diphenhydramine, as a potential drug that would reduce aversive memory recall (Papassotiropoulos et al. 2013). This was verified in a double-blind, placebo-controlled, cross-over study in which a single administration of diphenhydramine (50 mg) compared with placebo significantly reduced delayed recall of aversive, but not of positive or neutral, pictures.

Most disease-associated SNPs, however, occur in non-coding regions and their impact on disease outcome is difficult to

Table 3 Studies applying systems pharmacology approaches to brain diseases

Disease/condition	Organism/model	Tissue	Input	Similarity metric	Key findings	Validation	Measure of reliability/other prioritization methods	Public resources used	Ref
Alzheimer's disease (AD)	Human	Hippocampus	DEGs—top 500 up-regulated and top 500 down-regulated genes based on fold change (unadjusted P value ≤ 0.05). Calculated 3 different ways (Limma, Limma-ChiDir, and mAP-KL) for 5 independent studies (all hippocampus).	KS-like statistic. Correlation coefficient	Proposed 27 candidate drugs. Highlighted potential roles of PKC, HDAC, ARG, and GSK3 in mechanism of AD.	None	Negative scores; CMap (permuted results P value < 0.05); SPIEDw (results with significance value > 2); sscMap treatment set score normalized to unity with a tolerance of one false connection among all possible drugs ($P < 1/1309$, strictest parameterization); LINCSeq1000 (mean connectivity score across the four cell lines in which the perturbation connected most strongly to the query (best score $4 < -0.9$)). Systematically varied calculation of DEGs for input ($\times 5$ hippocampal datasets) and algorithms. Combined score across all.	GEO, CMap (build 2), sscMap, SPIEDw, LINCSeq1000, Enrichr, ChemBioServer, NetworkAnalyst, and mode of action by NeTwork Analysis (Mantra 2.0).	Siavelis et al. (2016)
AD	Human	Entorhinal cortex; hippocampus; middle temporal gyrus; posterior cingulate cortex; superior frontal gyrus; visual cortex.	DEGs—top 400 up-regulated and top 400 down-regulated genes based on fold change (FDR $< 1\%$).	KS-like statistic	Gene signature induced by thioridazine was similar with AD gene signatures in EC, HIP, MTG, and SFG. On the contrary, gene signatures induced by 2 histone deacetylase (HDAC) inhibitors vorinostat and trichostatin A were anticorrelated with AD gene signature in HIP and PC, which indicated that these drugs could possibly reverse the AD gene signature.	None	Positive scores (mechanistic insight). Negative scores (candidate pharmacotherapeutics).	GEO, DAVID, TransFind, CMap, and KEGG	Chen et al. (2013)
AD	Human	Hippocampus and cerebral cortex	DEGs—hippocampus: 40 DEGs (40 genes reported by Hata et al. 2001; top 20 up and top 20 down). Cerebral cortex: 25 genes with FC > 5 reported by Ricciarelli et al. 2004; 11 up and 14 down).	KS-like statistic	No genes in common between these two query signatures, but both yielded negative connectivity scores with the two independent instances of 4,5-diaminopthalimide (DAPH). This strengthened the candidacy of DAPH as a potential AD therapeutic.	In silico (literature: DAPH was found to reverse the formation of fibrils (a variety of DAPH analogs have been synthesized as potential treatments for AD).	Negative scores and permutation p value	CMap	Lamb et al. (2006)
AD/cognition enhancers	Human	Hippocampus and cerebral cortex	DEGs—hippocampus: 40 DEGs (40 genes reported by Hata et al. 2001; top 20 up and top 20 down). Cerebral cortex: 25 genes with FC > 5 reported by Ricciarelli et al. 2004; 11 up and 14 down).	KS-like statistic	No genes in common between these two query signatures, but both queries resulted in a common list of negative connections that were given higher confidence. They used an integrative chemoinformatics approach combining CMap with QSAR/VS hits to emphasize connections from the CMap that one would not choose otherwise.	In vitro (validated binding to 5HT ₁ 6R, also that raloxifene binds to 5HT ₁ 6R). In silico (literature: raloxifene given at a dose of 120 mg/day, but not 60 mg/day, led to reduced risk of cognitive impairment in postmenopausal women).	Negative scores (they mention statistically significant negative drugs but do not provide the threshold used.). Note: no summarization across multiple experiments for same compound (doses, timepoints, cell lines).	CMap	Hajjo et al. (2012)
Parkinson's disease (PD)	Human	Substantia nigra	DEGs—top 20, 50, 100, 200, or all (535) DE genes from integrated approach, 2 GEO datasets: FC > 1.5 and	KS-like statistic	Top 20 genes from their integrated approach for prioritizing genes outperformed the top 100, 200, 500, and all (536) DEGs	In vitro (neuroblastoma cell line). In silico (enrichment score)	Negative scores. Integrated approach (NOTE: simply using the DEGs from the GEO datasets performed as well as or better than their	CMap, GEO, ParkDB, OMIM, and CTD	Gao et al. (2014)

Table 3 (continued)

Disease/condition	Organism/model	Tissue	Input	Similarity metric	Key findings	Validation	Measure of reliability/other prioritization methods	Public resources used	Ref
Huntington's disease (HD)	Human	Caudate nucleus	FDR < 10%. ParkDB (human): consistent up- or down-regulation across different experiments ($P \leq 0.01$). DEGs—top 100 (absolute FC); 8 up-regulated and 92 down-regulated ($P < 0.05$)	KS-like statistic	for the input signature. Performance was measured by how many (out of the top 50) molecules returned by CMap from their approach were enriched with therapeutic molecules for PD. 1 candidate, alvespimycin (17-DMAG), was found to be neuroprotective in an in vitro rotenone model of PD. Using a gene signature for HD, CMap identified potential therapeutic agents with multiple modes of action and validated 2 (deferoxamine and chlorzoxazone) in vivo ameliorated neurodegeneration in flies expressing a mutant HTT fragment).	In vitro: luminescent caspase-activation assay of HTT-induced apoptosis in a PC12 cell line; 7/12 drugs with negative connectivity scores reduced HTT-induced apoptosis. Chlorzoxazone, copper sulphate, deferoxamine, febinaf, oligomycin and primidone did so in a dose-dependent manner. Chemicals with positive connectivity scores had little effect on caspase activation. In vitro: high-throughput recording of HTT103Q aggregation in PC12 cells using Cellomics imaging technology. Of the 7, deferoxamine and oligomycin, significantly altered the formation of HTT103Q-containing inclusion bodies. In vivo: deferoxamine and chlorzoxazone ameliorated neurodegeneration in flies expressing a mutant HTT fragment (a widely studied fruit fly model of mutant HTT toxicity) (Stefian et al. 2001)	Negative scores	CMap and ArrayExpress	Smalley et al. (2016)
Memory-modulating compounds	Human	Saliva	Intragenic SNPs associated with aversive memory recall	Not specified	They used genomic information related to aversive memory—a trait central to posttraumatic stress disorder—to identify several potential drug targets and compounds. In a subsequent pharmacological study with one of the identified	In vivo: a single administration of diphenhydramine (50 mg) compared with placebo significantly reduced delayed recall of aversive, but not of positive or neutral, pictures in a	Not specified	Ingenuity pathway analysis (IPA) (note: not public)	Papassotiropoulos et al. (2013)

Table 3 (continued)

Disease/condition	Organism/model	Tissue	Input	Similarity metric	Key findings	Validation	Measure of reliability/other prioritization methods	Public resources used	Ref
Motivation to exercise	Mouse (4 models of motivation to exercise and controls)	Striatum	DEGs—top 287 genes up-regulated. Top 235 genes down-regulated in selected lines vs. controls (FDR < 5%)	KS-like statistic	compounds, diphenhydramine, they found a drug-induced reduction of aversive memory. These findings indicate that genomic information can be used as a starting point for the identification of memory-modulating compounds. Mice from 4 lines selected for wheel running (and 4 non-selected lines) were allowed full access to a running wheel for 6 days. On day 7, half of the high runners and half of the low runners were blocked from wheel access and striatum taken at the time of maximum wheel running and submitted for RNA sequencing. LINC5 identified the protein kinase C δ inhibitor, rottlerin, the tyrosine kinase inhibitor, Lintifanib and the delta-opioid receptor antagonist 7-benzylidenalrexone as potential compounds that mimic the transcriptional signature of the increased motivation to run.	double-blind, placebo-controlled study in healthy volunteers	Positive scores (to mimic the high motivational state for exercise). Considered both compounds tested across all cell lines and those only tested in neuronal cell lines. No statistical p value mentioned.	DAVID and LINC5-L1000	Saul et al. (2017)
Schizophrenia, major depressive disorder, bipolar disorder, Alzheimer's disease, anxiety disorders, autistic spectrum disorders, and attention deficit hyperactivity disorder	Human	10 brain areas available in GTEx: anterior cingulate cortex (BA24), caudate (basal ganglia), cerebellar hemisphere, cerebellum, cortex, frontal cortex (BA9), hippocampus, hypothalamus, nucleus accumbens (basal ganglia), and putamen (basal ganglia)	GWAS summary statistics converted to transcriptomic profiles. Top K DEGs where K was varied for 50, 100, 250, and 500. Results were averaged across each K.	5 methods: KS-like statistic, Spearman correlation with all or with K differentially expressed genes and Pearson correlation with all or with K differentially expressed genes	They imputed transcriptomic profiles for 7 psychiatric conditions based on GWAS summary statistics and compared these to drug-induced changes in gene expression (CMap) to find possible treatments and found that the top 15 predicted compounds were enriched with known drug-indication pairs.	In silico	Negative scores, Permutation test (shuffled the disease-expression z -scores and compared them to drug transcriptomic profiles. Performed 100 permutations for each drug-disease pair and combined the distribution of ranks under the null across all drug-disease pairs, such that the null distribution was derived from 347,800 ranks under H_0).	CMap, GTEx, MetaXcan, KEGG, ClinicalTrials.gov, Medication Indication resource (MEDI), and Psychiatric Genomics Consortium (PGC)	So et al. (2017)
Morphine tolerance	Rat	Whole brain	DEGs—morphine-tolerant + saline versus morphine-tolerant + LPS; placebo-control + saline versus placebo-control + LPS; placebo-control + saline versus	KS-like statistic	Here, LINC5-L1000 gene knockdown and overexpression experiments were used to inform mechanism of action. Response to LPS was altered during morphine tolerance and indicated that VPS28	None	Genetic perturbation experiments only; positive scores; negative scores	LINC5-L1000 Query App (apps.lincscloud.org/query) (now deprecated. Instead use clue.io/11000-query)	Chang et al. (2017)

Table 3 (continued)

Disease/condition	Organism/model	Tissue	Input	Similarity metric	Key findings	Validation	Measure of reliability/other prioritization methods	Public resources used	Ref
Binge-like drinking (important risk factor for AUD)	Mouse (HDID-1 and HS/Npt controls)	Prefrontal cortex, nucleus accumbens core, nucleus accumbens shell, bed nucleus of the stria terminalis, basolateral amygdala, central nucleus of the amygdala, ventral tegmental area, and ventral striatum	morphine-tolerant + saline rats (no <i>p</i> value threshold reported) DEGs—top 100 up-regulated and top 100 down-regulated genes from each brain area (based on fold change, unadjusted $p < 0.05$). Differentially expressed landmark genes from each brain area (unadjusted $p < 0.05$) (landmark genes are those whose expression is directly measured in the L1000 assay)	KS-like statistic	may be one of the genes responsible for the alterations associated with morphine tolerance. Many anti-inflammatory compounds had highly negative connectivity scores across brain areas, providing additional evidence for a neuroimmune component in regulating ethanol intake. The top 2 candidates, terreic acid, and pergolide were behaviorally validated to decrease ethanol intake and blood alcohol levels.	In vivo: the top 2 candidates, terreic acid, and pergolide were behaviorally validated to decrease ethanol intake and blood alcohol levels.	Negative scores, integrated approach, and Sig_gmic tool (see text)	LINCS-L1000 and GEO	Ferguson et al. (2017)
Traumatic brain injury (TBI)	Rat	Perilesional cortex and thalamus	DEGs—top 4964 in perilesional cortex and top 1966 in thalamus (FDR < 5%)	Not reported	The study highlighted tubulins, <i>Nfe2l2</i> , <i>Nfya2</i> , and <i>S100a4</i> as target genes modulated by compounds with a high LINC connectivity score relative to the TBI-sig. Their data suggested that desmethylclomipramine, an active metabolite of the antidepressant clomipramine, is a promising TBI treatment candidate.	In silico: 2/11 top compounds had previously been investigated in epileptogenesis models in vivo.	Negative scores	LINCS-L1000, GEO, IPA, GSEA, and MsigDB	Lipponen et al. (2016)
Ischemic stroke	Rat	Brain (whole hemisphere)	DEGs—genes with $FC1 > 1.5$, $FC2 > 1.2$ and $RR > 0$. $FC1$: fold change between middle cerebral artery occlusion (MCAO) and Sham ($FC1$). $FC2$: fold change between MCAO and XST $RR = \frac{M_i - X_i}{M_i - C_i}$ where C_i , M_i , and X_i are the average expressions of gene <i>i</i> in control group, MCAO group, and XST treatment group, respectively.	KS-like statistic	This study sought to investigate the mechanism of action of a Chinese medicine called Xuesaitong injection (XST), a prescription drug made of Panax notoginseng that is used for treating stroke in China. They looked at positive scores. Inhibition of inflammatory response and coagulation were identified as the major mechanisms involved in the protective effects of XST.	None	Positive scores and permutation <i>p</i> values < 0.05	CMap	Wang et al. (2015)
Epilepsy	Human	Cerebellar cortex, temporal cortex, occipital cortex, hippocampus, thalamus, white	Epilepsy-associated co-expression module (M30)	Fischer's exact test	They used post-mortem human brain samples from healthy individuals from the UK Brain Expression Consortium (UKBEC) dataset to build gene co-expression networks	None. However, confirmed in vitro that VPA upregulates 51% of the M30 genes in neurons (FDR < 10%), replicating and strengthening the	Tested the overlap of M30 genes with the list of genes upregulated by a drug using one-tail Fischer's Exact Test (FET) (in order to prioritize drugs predicted to reverse the downregulation of M30	UK Brain Expression Consortium, Genotype-Tissue Expression (GTEx) project, STRING, WebCestalt,	Delahaye-Duriez et al. (2016)

Table 3 (continued)

Disease/condition	Organism/model	Tissue	Input	Similarity metric	Key findings	Validation	Measure of reliability/other prioritization methods	Public resources used	Ref
		matter, medulla, and putamen			(modules) and integrated modules with whole-exome-sequencing (WES) studies data of rare de novo mutations in those with epileptic encephalopathy (EE). A single module was selected: M30. The M30 genes' functional expression for 3 epilepsies suggested <i>downregulation</i> of the network as a common mechanism. They used CMap to identify drugs that could up-regulate the M30 genes. Valproic acid (VPA), a widely used antiepileptic drug, was the drug most significantly predicted to up-regulate the genes in M30 (toward health).	VPA-signature in the cancer cell lines from CMap.	genes observed in epileptic hippocampi. BH-corrected <i>p</i> values for multiple hypotheses testing. Included only 152 drugs with ≥ 10 DEGs (FDR < 10%). No summarization across cell lines, doses, and timepoints for each drug	GeneMANIA, Hippic, iRefWeb, HUGO Gene Nomenclature Committee database, and CMap	
Epilepsy	Mouse (pilocarpine-induced chronic epilepsy)	Hippocampus	DEGs—FDR < 0.05 and fold change ≥ 2 (929 up, 1164 down)	KS-like statistic (used query app on lincseloud.org)	The authors queried the LINCSEL1000 database with an epilepsy signature consisting of the top DEGs between the hippocampus of a model of epileptic mice and control mice. They identified 123 compounds with negative scores beyond a chosen threshold (mean of best 4 ≤ -85). These 123 compounds were enriched with compounds known to have antiepileptic effects. Despite a diverse set of mechanisms of action, these compounds targeted similar biological pathways and were better at reversing pathways affected by epilepsy than common antiepileptic compounds. After filtering for practical exclusion criteria, 36 compounds remained. Of these, sitagliptin was confirmed to have antiepileptic activity <i>in vivo</i> .	In silico (the 123 compounds with LINCSEL mean connectivity score threshold of -85 or less were 6-fold more enriched with antiepileptic drugs (7/123) than the drugs in the LINCSEL database as a whole (203/19,767)). <i>In vivo</i> (Sitagliptin produced a dose-dependent reduction in seizure scores) in the 6 Hz psychomotor seizure mouse model of pharmacoresistant epilepsy)	Negative scores. Inclusion criterion: LINCSEL mean connectivity score threshold of -85 (123 compounds). Exclusion criteria: toxicity, parenteral route of administration, lack of animal or human dosage data, or BBB-impermeability (36 compounds remained). This was the only study on the table that used any practical considerations to filter compounds. No published evidence of BBB-impermeability for any of the drugs.	LINCSEL1000, drug-set enrichment analysis (DSEA), and gene-set enrichment analysis (GSEA)	Mirza et al. (2017)
Nerve regeneration	Rat	Dorsal root ganglion neurons. 13 separate studies: a total of 382 gene expression datasets (microarray) related to nerve injury	(1) PPI network consisting of 280 genes. (2) Regeneration-associated co-expression modules	KS-like statistic	The authors identified a transcriptional program observed after peripheral, but not central, nerve injury. They used the regeneration-associated modules to query CMap (2 different inputs) and tested the top 3 candidates that emerged from the intersection of the 2 queries (ambroxol, lasalocid, and disulfiram). Only ambroxol	In vitro: ambroxol, lasalocid, and disulfiram were tested, but only ambroxol enhanced axonal outgrowth of DRG neurons. <i>In vivo</i> : ambroxol increased optic nerve (ON) regeneration in mice after a crush injury, albeit a modest improvement.	Positive scores and permutation <i>p</i> value	CMap, GEO, PubMapix, DAVID, JANSFAC, TRANSFAC, STRING, and ENCODE	Chandran et al. (2016)

Table 3 (continued)

Disease/condition	Organism/model	Tissue	Input	Similarity metric	Key findings	Validation	Measure of reliability/other prioritization methods	Public resources used	Ref
CNS injury	Human MCF7 breast adenocarcinoma cells	NA	DEGs—absolute fold change ≥ 1.5 , $p < 0.05$ (10 up and 12 down); 6 h treatment of F05 (5 μM); a compound the group previously identified to promote neurite growth in vitro and in vivo) vs. vehicle (DMSO, 0.05%)	KS-like statistic	enhanced axonal outgrowth of DRG neurons in vitro and it also increased optic nerve (ON) regeneration in mice after a crush injury, albeit a modest improvement. The authors used the transcriptional signature of a compound known to induce neurite growth (F05) as a “seed” to identify other compounds with this same property. Remarkably, despite no chemical similarity to F05, a group of piperazine phenothiazine antipsychotics had similar effects on gene expression and were found to promote neurite growth in vitro at least partially through antagonism of calmodulin signaling, independent of dopamine receptor antagonism.	In vitro: $\frac{3}{4}$ piperazine phenothiazine antipsychotics (but none of the other classes of antipsychotics) significantly enhanced neurite growth of dissociated hippocampal neurons and rat retinal ganglion cells on a substrate of a mixture of chondroitin sulfate proteoglycans (CSPGs) (glial scar proteins)	Positive scores	CMap	Johnstone et al. (2012)
CNS injury/-neurodegeneration	Mouse (Ascl1-E-GFP ^{Cre} transgenic reporter mouse line)	Subventricular zone (SVZ) of the dentate gyrus microdomains Region-specific neural stem cells (NSCs) and their immediate progeny. Transient amplifying cells (TAPs)	DEGs—1.8-fold change and FDR $< 5\%$	Pearson correlation (via SPIED web-based tool)	This study aimed to identify compounds that could direct germinal activity in the subventricular zone (SVZ), which would have therapeutic potential in nerve injury/neurodegenerative or demyelination diseases. They examined NSC lineages in the SVZ microdomains (dorsal versus ventral/lateral) and identified small molecules that direct the fate of these cells toward neurogenic as opposed to oligodendrocyte lineages. LY-294002, an inhibitor of PI3K/Akt, induces transcriptional changes that promote oligodendrogenesis. AR-A014418, an inhibitor of GSK3 β , induces transcriptional changes that promote rejuvination of the adult SVZ.	In vivo: infusion of AR-A014418 or CHR99021 in the adult (P90) dSVZ lateral ventricle dramatically stimulated the germinal activity of the adult dSVZ. CHR99021, a GSK3 β inhibitor, showed regenerative potential in a neuro-pathological context in a model of premature injury that leads to diffuse oligodendroglial and neuronal loss throughout the cortex	Positive scores	CMap, GeneGO Metacore for Process Networks, and SPEDw	Azim et al. (2017)
Down Syndrome (DS)	Human and mouse (3 models of DS; Dn16, Ts65Dn and Ts1Cje and controls)	Human: second trimester amniocytes; induced pluripotent stem cells (iPSCs); neurons derived from iPSCs; post-mortem human fetal cerebellum and	DEGs—BH-FDR $< 5\%$ and 20%. Because of the limited number of differentially regulated genes at FDR 20%, they used the top 1% up- and down-regulated genes	KS-like statistic	The authors used a human/mouse integrated approach to identify 17 high priority molecules predicted to reverse pathway changes in both human cells and mouse models. They tested the effects of apigenin, one of the molecules predicted by the CMap to treat dysregulated pathways in	None—currently undergoing in vitro and in vivo testing (not yet published)	Negative scores, Threshold: -0.7 or less. Integrated approach (multiple inputs)	GSEA, DAVID, IPA, GEO, and CMap	Guedj et al. (2016)

Table 3 (continued)

Disease/ condition	Organism/ model	Tissue	Input	Similarity metric	Key findings	Validation	Measure of reliability/ other prioritization methods	Public resources used	Ref
		cerebrum. Mouse: developing forebrain (E15.5)			DS, on human amniocytes derived from fetuses with DS and on the Ts1Cje mouse model of DS. Apigenin treatment reduced oxidative stress in DS amniocytes and improved some aspects of brain morphogenesis, gene expression and postnatal behavior in the Ts1Cje mouse model (manuscript in preparation).				
Down syndrome	Human	Second trimester amniotic fluid	DEGs—FDR < 5% (414 probes individually differentially expressed between trisomy 21 and controls)	KS-like statistic	This is one of the first studies to demonstrate that transcriptional profiling of RNA in uncultured amniotic fluid provides molecular insights into developmental disorders in the living human fetus. They found 4 compounds with average connectivity scores > 0.7 (indicating a high correlation with the DS molecular signature), and 9 compounds with average connectivity scores less than - 0.7 (indicating a high negative correlation) (NSC-5255229, celestrol, calmidazolium, NSC-5109870, dimethyl/oxalyglycine, NSC-5213008, verapamil, HC toxin, and felodipine). The 4 compounds that most mimic the DS phenotype were related to potassium and calcium signaling or oxidation which further supports the importance of oxidative stress and ion transport as functional classes involved in DS.	None	Positive scores (mechanistic insight), Negative scores (candidate pharmacotherapeutics)	GSEA, DAVID, and CMap (build 1.0)	Slonim et al. (2009)

causally link to gene expression, although this is an active area of research and databases exist trying to relate SNPs with gene expression in a variety of tissues including brain (e.g., Genotype-Tissue Expression Project; <https://www.gtexportal.org/home/>). One study approached this problem in an innovative way for psychiatric illnesses and could be considered a hybrid approach because the authors inferred gene expression data from genotype data (So et al. 2017). The approach relied on an algorithm called *MetaXcan* (Barbeira et al. 2016), which incorporated GTEx data to build statistical models for predicting expression levels from SNPs in a reference transcriptome dataset, and these prediction models were used to impute the expression *z*-scores (i.e., *z*-statistics derived from association tests of expression changes with disease status) based on GWAS summary statistics. Transcriptome profiles were imputed for seven psychiatric conditions based on GWAS summary statistics and compared with drug-induced changes in gene expression using CMap to identify potential treatment candidates. Novel compounds were not tested; however, it was promising that the top 15 predicted compounds for some of the psychiatric disorders were enriched with known and predicted psychiatric medications according to several drug-disease indication measures (Anatomical Therapeutic Chemical (ATC) codes, ClinicalTrials.gov, Medication Indication (MEDI) resource).

Once the input genomic signature is defined, it can be compared to a database of drug signatures. Most of the studies in Table 3 (13/20) use the original CMap database. The benefit of CMap is that it is smaller and simpler to perform statistics to assess a connectivity score's reliability. However, the trade-off is fewer drugs and cell lines, the latter of which is especially important for brain diseases because CMap contains no brain cell lines, whereas LINCS-L1000 contains two brain cell lines with considerable data, NEU and NPC. Unfortunately, at this time, these cell lines are not included in the implementation on their query app at clue.io; however, the LINCS-L1000 datasets can also be downloaded from Gene Expression Omnibus (GEO) (accession numbers GSE70138 and GSE92742). Efforts are underway to include more brain cell lines in the LINCS-L1000 database to facilitate its relevance to brain diseases (RDM, personal communication).

To compare the disease and drug signatures, the KS-like statistic (as described by (Lamb et al. 2006; Subramanian et al. 2017) is the most frequently used similarity metric, although several studies also use Spearman or Pearson correlation coefficients (Azim et al. 2017; Siavelis et al. 2016; So et al. 2017) or Fischer's exact test (Delahaye-Duriez et al. 2016). Most studies operate under the transcriptional "reversal hypothesis," which assumes that drugs with negative connectivity scores (i.e., with gene expression signatures that revert the disease's effects on gene expression to the control state) would ameliorate disease phenotype. Five of the 20 studies outlined in Table 3 have functionally validated this hypothesis, in that the candidate compounds ameliorated some disease

phenotype when tested behaviorally (though none have confirmed that the beneficial effects of the compound were due to the restoration of gene expression to the "normal" state) (Chandran et al. 2016; Ferguson et al. 2017; Mirza et al. 2017; Papassotiropoulos et al. 2013; Smalley et al. 2016).

These studies provide a functional rationale for prioritizing negatively-scoring compounds, i.e., those that have opposing effects on gene expression associated with the disease state. However, in addition to reflecting gene expression changes that drive the disease or represent deleterious aspects of a disease state, the differentially expressed genes between disease and healthy samples could also reflect protective homeostatic compensations within the system. Because some of the differentially expressed genes might be beneficial, it is reasonable to also consider drugs with high positive connectivity scores.

The rationale for the reversal hypothesis was tested directly utilizing a gene expression signature comprised of the top 100 differentially expressed genes identified in Huntington's disease (HD). Data were obtained from the caudate nucleus from disease vs. sex- and age-matched human controls followed by CMap query (Smalley et al. 2016). The top 12 positive and negative scoring compounds were tested in *in vitro* caspase-activation assays to assess the degree to which they modulated mutant huntingtin (HTT)-induced apoptosis in a PC12 cells. None of the positive scoring compounds affected caspase activity, while 7/12 negative scoring compounds decreased caspase activity, two of which had neuroprotective effects *in vivo* in a *Drosophila* model of HD. This outcome supports the "reversal hypothesis." However, because the caspase activity was approaching 100% (i.e., a ceiling effect), the ability to observe increased caspase activity precluded experimental outcomes predicted from positive scores that might mimic/worsen disease phenotypes (the converse of the reversal hypothesis).

One study does provide *in vivo* validation of the converse of the reversal hypothesis: that drugs with positive scores (i.e., with gene expression signatures that are similar the disease's effects on gene expression) would mimic the state of interest. Azim et al. (2017) sought to identify small molecules to mobilize endogenous stem cells and direct their fate as a therapy for neurodegenerative and demyelinating disorders (Azim et al. 2017). These studies used the transcriptional signatures of neural stem cells (NSCs) in the ventral/lateral subventricular zone (SVZ) of the dentate gyrus which give rise to interneurons of the olfactory bulb and cortical areas, and of NSCs in the dorsal SVZ which give rise to glutamatergic neurons and oligodendrocytes. The authors prioritized positively-scoring compounds with the hopes that that would reproduce the lineage-specific transcriptional signatures. Indeed, the most promising candidates, LY-294002, an inhibitor of PI3K/Akt, promoted development of oligodendrocytes, and AR-A014418, an inhibitor of GSK3 β , rejuvenated the NSC lineage. Furthermore, another GSK3 β inhibitor

promoted regeneration in a mouse model of hypoxic brain injury, by recruiting new oligodendrocytes and glutamatergic neurons into the cortex.

In addition to gene expression signatures of drug perturbation, the LINCS-L1000 database also catalogs gene expression response to genetic perturbation. One study utilized this resource and compared the input genomic signature to those of gene knockdown or overexpression in LINCS-L1000 to gain mechanistic insight into how morphine tolerance alters response to lipopolysaccharide (LPS) and found that VPS28 may be one of the genes responsible for the alterations associated with morphine tolerance (Chang et al. 2017). In addition to looking at the negatively correlated drugs for treatment candidates, several studies also analyzed the positively correlated drugs for mechanistic insight into the disease, as these would be predicted to produce similar effects on gene expression and mimic or worsen disease phenotype (Chen et al. 2013; Slonim et al. 2009).

Challenges and future directions

The CMap and LINCS-L1000 databases contain multiple experiments for the same compound. It is clear that the compound's effects on gene expression are greatly affected by variables such as cell line, dose, and time point at which gene expression is assayed (Chen et al. 2017). Some researchers make no attempt to summarize across cell lines, doses, and time points to attain a composite compound-level view, which could lead to spurious results, particularly if the cell line is vastly different from the cellular makeup of the tissue used to generate the genomic signature used as the input query. For these reasons, we propose that using multiple expression datasets, algorithm parameter settings, and methods for prioritizing compounds (as taken by Ferguson et al. 2017; Gao et al. 2014; Guedj et al. 2016; Siavelis et al. 2016) are critical to identify an effective drug candidate, at least until proper gold standard datasets exist with which to benchmark the optimal settings (see below).

An important limitation of *in silico* gene mapping approaches is that they rely on comparisons of brain gene expression data to gene expression data from cell culture and are therefore constrained by the same limitations of any *in vitro* system. Brain gene expression is a complex combination of direct and indirect expression changes occurring in multiple cell types and brain regions. Even if brain-relevant cell lines were included in the expression profiles of LINCS-L1000 or other databases of drug-related transcriptomes, it is unclear how relevant *in vitro* results are to the biology of an intact organism, which is why *in vivo* experimental validation is critical. Only 6 of the 20 studies for brain disease listed in Table 3 performed *in vivo* validation of the proposed pharmaceutical candidates (Azim et al. 2017; Chandran et al. 2016; Ferguson et al. 2017; Mirza et al. 2017; Papassotiropoulos et al. 2013; Smalley et al. 2016), and none directly tested the

underlying assumption of *in silico* connectivity mapping. Specifically, it is important to address the following question: if a candidate compound is effective in treating a given disease phenotype, was it the result of a reversal in expression of disease-related genes by the compound? This is difficult to assess given the complexity of the regulation of gene expression. Parameters such as drug dose and treatment times are critical for determining meaningful gene expression changes. Therefore, a range of doses and time points would need to be measured, and although the cost of whole genome sequencing is decreasing, to do this with the required samples sizes would be cost prohibitive. In addition to L1000 technology, the development of less expensive sequencing techniques, such as TagSeq, improve feasibility to test this hypothesis which will provide important mechanistic insight into *in silico* gene mapping approaches (Lohman et al. 2016; Meyer et al. 2011).

Each of the six studies discussed in the previous paragraph used a different approach to identify a candidate compound that ameliorated disease phenotype when tested (Azim et al. 2017; Chandran et al. 2016; Ferguson et al. 2017; Mirza et al. 2017; Papassotiropoulos et al. 2013; Smalley et al. 2016), and it is critical to identify the approach(s) with the greatest predictive accuracy. In other words, which choices at each of the main steps outlined above are the most likely to identify compounds that actually ameliorate the disease state? It is currently difficult to address this question because of the low-throughput nature of behavioral testing and the non-existence of gold standard data with which to benchmark various approaches.

A benchmark approach requires a gold standard dataset comprised of two components from the same population: (1) gene expression and/or genotyping data. Ideally, gene expression data would be obtained from multiple brain areas, cell types (single cell or cell-type transcriptomes), and tissue types (peripheral blood mononuclear cells (PBMCs), liver, gut microbiome, etc.) and (2) drug response. This should be the same measurement for each drug and there would ideally be a large range of drug effects. This continuous variable would lend itself to correlation analysis (rather than a binary measure of 0: drug was ineffective and 1: drug was effective). Drugs that are known to affect the phenotype (true positives) and drugs that are known to not effect phenotype (true negatives) should be present to assess how well the approach can discriminate (assign high scores to the true positives and poor scores to the true negatives). A benchmarking test-case scenario using this ideal gold standard dataset would systematically vary the input, algorithm, and prioritization scoring choice and assess the outputs for their predictability (Table 4).

One caveat to this benchmarking strategy outlined here, is that it is unreasonable to assume that all compounds with therapeutic potential would be identified by *in silico* gene mapping. The best way to evaluate these approaches would be to take a heuristic testing strategy and select a few compounds nominated from various combinations at each of the

Table 4 Benchmarking test case

1. Input	2. Similarity metric	3. Prioritization
Tissue: whole brain, brain regions, cell type-specific transcriptome, and single cell transcriptome Genes: SNPs, differentially expressed (top 50, 100 → 500); co-expression modules	Correlation, enrichment (hypergeometric, Fischer's, modified KS statistic), and pattern matching	Statistical methods; negative score vs. absolute value of score; threshold (for example, −90 cutoff); median; combination of above

three steps to test behaviorally, but as mentioned previously, behavioral testing is low throughput and this would be resource intensive.

As discussed before, the affected tissue (brain) is not available for testing until post-mortem, which certainly poses a problem if computational approaches that rely on gene expression measures are to be incorporated into drug repurposing/personalized medicine endeavors for brain diseases. Moreover, analysis of postmortem brain expression is plagued with the “chicken and the egg” conundrum. Meaning that it is impossible to know if the observed gene expression changes are the cause or the effect (due to years of alcohol use, for example) of the disease. This is one reason why animal models are key in studying brain diseases, and using animal models with high predictive validity for selecting therapeutic compounds is one way around this problem. Another option would be to identify a surrogate for brain gene expression, and this is an active area of research. Great hope has arrived with the discovery of inducible pluripotent stem cells (iPSCs) that can be differentiated into various neuronal types (Takahashi and Yamanaka 2006). There are also methods that skip the induced pluripotent steps allowing direct conversion into functional neurons, called induced neurons (or iNs; for review, see Drouin-Ouellet et al. 2017). Although these cell models hold promise for improving treatment of psychiatric diseases (Oni et al. 2016; Stern et al. 2017), the protocols are long and tedious to produce adult-like neurons and the relevance to *in silico* gene mapping remains unexplored. Another surrogate for brain gene expression could be in peripherally accessible cell types, like PBMCs, especially considering the impressive evidence for immune involvement in AUD (for reviews, see Crews et al. 2017; de Timary et al. 2017; Mayfield and Harris 2017). Another option that does not require access to brain tissue is imputing gene expression from GWAS summary data as discussed above (So et al. 2017). However, the latter approach has yet to be validated *in vivo* and will likely improve as the databases used to make the imputations improve, for example, by increasing samples in GTEx to better detect eQTLs. The GTEx Consortium plans to include up to 1000 donors in the final data release and collect complementary molecular data on subsets of samples, including epigenetic and protein data (GTEx Consortium et al. 2017). Using GTEx data to impute transcriptomes for diverse groups of

people should be approached with caution, as the donors are currently 83.7% European American and 15.1% African American (with the v7 Release) (GTEx Consortium et al. 2017).

Conclusion

The benefits of using computational strategies to transition to a more molecularly informed healthcare system are numerous. For example, diagnoses could be more precise and treatments more successful. Patients diagnosed with the same disease often represent a heterogeneous mixture of different underlying disorders, because there are numerous molecular disruptions that could lead to similar clinical presentations. This is especially true for AUD and other brain diseases, where a molecular readout of the affected organ is limited. It is no surprise, then, that the standard treatments fail for many because of incomplete knowledge regarding the underlying cause of a patient's disruptive symptoms. As we become more advanced in our ability to construct and interpret a molecular signature underlying disease symptoms, healthcare will advance toward personalized medicine, where each patient is treated to his or her individual profile.

The systems pharmacology approaches discussed in this Review have two main beneficial outcomes that should be considered independently. The first is an effective treatment and the other is mechanistic insight. It might be that the effectiveness of a compound is understood before its mechanism of action. However, progressing promising pharmaceutical treatment should not wait for the full understanding of the mechanism, as the mechanism underlying some of the most long-standing and successful treatments in medicine are still poorly understood (Letai 2017). In fact, as suggested by (Hajjo et al. 2012), one of the main benefits of this approach is to identify potentially therapeutic compounds without necessarily understanding the underlying target-specific mechanism.

Much hope has been placed on information contained within large genomic datasets and network approaches to drive clinical treatment toward personalized medicine and revolutionize healthcare. And, indeed, bioinformatics approaches have shown some success for identifying novel treatments for brain diseases. However, this research is still in its infancy,

and many questions remain to be answered if these high expectations are to be met. Here, we have proposed the steps required for in silico gene mapping for the purpose of drug discovery and repurposing, reviewed state-of-the-art applications of these approaches to brain diseases, and highlighted some of the critical challenges facing the field. Success relies on the integration of enormous amounts of sequence and phenotype data from public and private sector sources. Ultimately, it will take a collaborative effort from academia, industry and government to advance drug development and repurposing for AUD (Litten et al. 2014).

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